

Research Article



Journal of Biomedical and Pharmaceutical Insights (JBPI)

e-ISSN: Applied For

Volume 1 | Issue 1 | 2026 | Pages 107-122.

DOI: To be assigned

<https://jbpijournal.org>

***IN-VITRO* CHARACTERIZATION AND EVALUATION OF
GASTRORETENTIVE FLOATING HBS CAPSULES OF GLICLAZIDE
FOR SUSTAINED DRUG RELEASE**

Radheshyam Samanta ^{1*}, Tamal Deb Goswami ¹, Hrushikesh Behera ², Ranjit Kumar Sutar ², Ashok Kumar Sethi ³, Pravati Deo ³, Bhabatarini Jena ⁴

¹Department of Pharmaceutics, Kalinga Institute of Pharmaceutical Sciences, Biruan, Balasore, 756060, Odisha, India

²Department of Pharmacology, Kalinga Institute of Pharmaceutical Sciences, Biruan, Balasore, 756060, Odisha, India

³Department of Pharmaceutical Chemistry, Kalinga Institute of Pharmaceutical Sciences, Biruan, Balasore, 756060, Odisha, India

⁴Department of Pharmacy, Kalinga Institute of Pharmaceutical Sciences, Biruan, Balasore, 756060, Odisha, India

ABSTRACT

The present research work attempted to formulation, characterization and evaluation of gastroretentive floating HBS (Hydrodynamically Balance System) capsules of gliclazide for sustain drug release. These floating HBS capsules of gliclazide were formulated by using two suitable polymer cationic charge chitosan and anionic charge xanthan gum with the help of physical blending and filled in capsules. Different evaluation parameters like their weight uniformity, drug content uniformity, swelling index of all formulation showing good results to develop the system. *In-vitro* floatation and in-vitro drug release studies also prove that all these formulation specially formulation GFC4 showing better drug release which is more than 10 hours due to suitable ratio using polymeric blend as well as formation of polyelectrolyte complex between two opposite charge polymers. It is also confirmed that there are no more incompatibility produce between drug and polymers by the help of FTIR also help to develop

the system. So, these types of gastroretentive floating HBS capsules will be very effective for gastroretentive sustain drug release.

Keywords: Gastroretentive, Floating, HBS, FTIR

This article is licensed under a Creative Commons Attribution-NonCommercial 4.0 International License. Copyright © 2026 The Author(s).

Received: 23-05-2026
Revised: 02-06-2026
Accepted: 06-06-2026
Published : 11-06-2026

Corresponding Author : Dr. Radheshyam Samanta
Email Id : radheshyamsamanta93@gmail.com
ORCID ID : 0009-0000-8145-8563

INTRODUCTION

The oral control release delivery of drug has been recently increasing interest in pharmaceutical department to obtain improved therapeutic advantages such as ease of dosing administration, patient compliance and easy to formulate ⁽¹⁾. Previously, several oral control release drug delivery system being designed and developed, including floating systems that causes buoyancy in gastric fluid help to improve the therapeutical response ⁽²⁾. Many approaches have been mention for gastroretentive sustain release drug delivery system through floatation ⁽³⁾, bio or mucoadhesion ⁽⁴⁾, sedimentation ⁽⁵⁾, unfoldable, expandable, or swellable systems ⁽⁵⁾, super porous hydrogel systems ⁽⁶⁾, magnetic systems ⁽⁷⁾, etc. Every approach has its own demerits. For example, swelling and expanding systems showing a hazard of fixed retention in the located site and muco or bioadhesive systems may cause in irritation of mucous layer because of high-localized concentration of the incorporated drugs ⁽⁶⁾, which produce serious complication for the patient. Among various gastroretentive drug delivery systems floating delivery of drugs represent the most effective and rational protection against early and rapid times of gastric emptying. Floating drug delivery system are designed to be remained floated on the gastric content due to its lower bulk density differentiated to that of the aqueous medium, thus retained in the upper region of GI tract for several hours and the drug is slowly release at a desired rate ^(4, 5). This results in an increased gastric residence time and a improve control of

the fluctuation in plasma drug concentration and however increase the bioavailability. Floating delivery of drug is of particular interest for drugs which ⁽⁶⁻⁸⁾ (a) act locally in the upper GI region (b) are primarily absorbed in the stomach (c) are poorly soluble at an high alkaline pH (d) have a narrow window of absorption and (e) are unstable in the intestinal or colonic environment. The main needed for floating delivery of drugs including that it should release contents slowly to serve as a reservoir, it should maintain specific gravity lower than gastric contents (1.004 – 1.01 gm/cm³), it should form a cohesive gel barrier ⁽⁸⁾. Floatation has been achieved with the preparation of low density solid systems e.g. inclusion of sponges, highly porous systems ⁽⁵⁻⁶⁾ which decrease in density upon contact with gastric fluid contents based on the expansion of swelling agents ⁽⁷⁾ or carbon dioxide generation ⁽⁸⁾. In addition, the inherent low density can be provided by the entrapment of air (e.g. hollow chambers) ⁽⁶⁾ or by the incorporation of low density materials (e.g. fatty materials or oils, or foam materials) ⁽⁵⁾.

Many attempts have been design to retain the floating dosage form in the stomach as a way of increasing the Gastric Residence Time (GRT) due to their low bulk density than gastric fluid and so remain floated in the stomach without altering the Gastric Emptying Rate (GER) for a prolong periods of time. When the system is floating on the gastric contents the drug is released sustain over prolong periods of time from this dosage form. After release of drug the intact system is emptied from the stomach which increased gastric residence time (GRT) and a improve control of fluctuations in plasma drug concentration. Although besides a minimal gastric content required to allow the proper achievement of the floating retention principle, a minimal level of floating force (A) is also needed to keep the dosage form reliably floated on the gastric fluid. To measuring the floating force kinetics a novel instruments for determination of resultant weight has been previously reported. The apparatus use by measuring continuously the force equivalent to A (as a function of time) that is required to maintain the submerged object. The object floats well if A is on the higher positive side. This apparatus helps in optimizing floating drug delivery systems with respect to stability and durability of floating forces produced in order to prevent the drawbacks of unforeseeable intragastric floating capability variations ⁽⁶⁻⁸⁾:

$$A = A_{\text{buoyancy}} - A_{\text{gravity}} = (d_f - d_s) v g$$

(Where, A= total vertical force, d_f = fluid density, d_s = object density, v = volume and g = acceleration due to gravity)

Based on the mechanism of floatation, two different technologies available, i.e. non-

effervescent systems and effervescent systems are used in the development of floating drug delivery systems⁽³⁾. Effervescent systems having gas generating systems including Intragastric single-layered floating tablets, Intragastric bi-layered floating tablets, Multiple-unit type of floating pills and Volatile liquid or vacuum containing systems including Intra gastric osmotically controlled floating delivery systems. Gas filled floating delivery systems. Non-effervescent systems contain hydrodynamically balanced systems (HBS), Microballoons (Hollow microspheres) and Floating beads^(3, 5, 7). Among these floating systems, HBS (hydrodynamically balanced systems) have showing a lot of importance in recent days to increase gastroretention, which improve absorption of drugs especially those are absorbed from stomach and small intestine or drugs such as weak bases, which dissolve better in the acid environment of the stomach⁽⁹⁾. These systems contain drugs with gel-forming hydrocolloids meant to remain buoyant for several hours in the stomach content. On contact with the gastric fluid, these systems form a water colloidal gel barrier around their surface and maintain a bulk density less than 1. They are mainly single-unit dosage forms, and usually composed of one or more gel-forming hydrophilic polymeric substances and an active pharmaceutical ingredient. Hydroxypropyl Methylcellulose (HPMC), Hydroxethyl Cellulose (HEC), Hydroxypropyl Cellulose (HPC), sodium Carboxymethyl Cellulose (NaCMC), polycarbophil, polyacrylate, polystyrene, Polyethylene Oxide (PEO), agar, carrageenans or alginic acid are commonly used excipients to develop these systems. The polymer is mixed with drugs and usually administered in the HBS capsules⁽¹⁰⁾. When the capsule containing drug-hydrocolloid mixture is exposed to gastric fluid, the capsule shell dissolves and the mixture swells to form a gelatinous barrier. This imparts buoyancy in gastric juice for a long period due to its continuous erosion of the surface, which allows water penetration to the inner layers maintaining surface hydration and buoyancy to the dosage form⁽¹¹⁾.

In this present work, gliclazide gastroretentive floating HBS capsules prepared by chitosan and xanthan gum by physical blending method was developed. These gliclazide floating HBS capsules were evaluated their uniformity of weight, swelling activity, drug content uniformity, *in-vitro* floatation; *in-vitro* drug release with kinetics and FTIR studies was performed.

MATERIALS AND METHODS

Materials

Gliclazide was a gift sample from Ajanta Pharma. Pvt. Ltd., India

Other materials were purchased from different sources like

Chitosan and xanthan gum: Sigma Aldrich, USA.

Talc, lactose, magnesium stearate: Loba Chemie Pvt. Ltd., India

All other chemicals and reagents used were of analytical grade.

Methodology

Preparation of gliclazide gastroretentive floating HBS capsule

Generally gliclazide floating HBS capsules was prepared by simple mixing (Blending) of gliclazide with polymers like chitosan and xanthan gum with their required quantities then mix other excipients like lactose as a diluents, talc as a glidant and magnesium stearate as a lubricant with this drug-polymer mixture for some time. Finally the mixture of gliclazide, chitosan, xanthan gum, lactose, talc and magnesium stearate carefully filled by 1 size hard geelatin capsules with special precaution for maintaining the drug content and weight uniformity. The quantity of different gliclazide floating HBS capsule is given in the following Table 1 (12-14, 16, 19).

Table 1: Formula of different ingredients used to prepare gliclazide gastroretentive floating HBS capsules

Formulation Code	Ingredients					
	Gliclazide (mg)	Chitosan (mg)	Xanthan gum (mg)	Lactose (mg)	Talc (mg)	Magnesium stearate (mg)
GFC1	100	25	125	138	2	10
GFC2	100	50	100	138	2	10
GFC3	100	75	75	138	2	10
GFC4	100	100	50	138	2	10
GFC5	100	125	25	138	2	10

Evaluation of uniformity of weight

For estimation of weight uniformity of prepared capsules by taking capsule of 20 numbers in each formulation and weight suitably by using analytical electrical balance that should help to determining the average or mean weight of these capsules. Then calculate the weight variation or % coefficient variation by using this formula (14-16).

$$\% \text{ coefficient variation} = \text{standard deviation} / \text{mean or average weight} \times 100$$

Evaluation of Swelling Index

The produced capsules were immersed in an excess amount of simulated gastric fluid at a temperature of 37°C for a specified duration. Subsequently, a time interval the capsules were

promptly extracted and measured using an analytical balance. The swelling percentage of the beads was determined using the following formula^(17, 18, 20).

$$\% \text{ swelling} = (\text{swollen beads weight} - \text{dry beads weight}) / (\text{dry beads weight}) \times 100$$

Evaluation of drug content uniformity

The drug content uniformity of floating capsules was determined by using glass beaker in a magnetic stirrer. Emptying 10 capsules from each formulation were placed in the beaker containing 500 ml simulated gastric fluid (pH 1.2) maintained at 37 ± 0.5 °C for 1 h and speed of this beaker containing magnetic stirrer at 500 rpm. After one hour sample was withdrawn and check spectrophotometrically at 230 nm after filtration with a suitable filter papers^(12, 14, 17, 21 - 22).

Evaluation of *in-vitro* floatation

For evaluating of *in-vitro* floating ability of gliclazide floating HBS capsules, immersed the one capsule from each formulation in 500 ml of simulated gastric fluid (pH 1.2) in a galas beaker at maintaining 37 ± 0.5 °C temperatures for required time. Then visually notice the capsule was float/buoyancy on simulated gastric fluid (pH 1.2) in particular duration of time and noted the time periods for express about the floating periods of these capsules^(14, 17, 23)

Evaluation of *in-vitro* drug release study

In-vitro drug release data of gliclazide floating HBS capsules in simulated gastric fluid (pH 1.2) was determined by using dissolution instrument (USP type II) with maintaining speed 50 rpm at 37 ± 0.5 ° temperatures. At first one capsule of gliclazide was transfer to the simulated gastric fluid (pH 1.2) containing basket rotational dissolution instrument then at the intermediate time interval 5 ml of sample withdraw from the dissolution medium and same quantity of fresh phosphate buffer (pH- 7.4) was added for maintaining sink condition. Now this withdraw sample from the dissolution medium was diluted and filtered by using Whatmann^R filter paper and estimate the amount of gliclazide release using a spectrophotometer (Double beam UV-VIS spectrophotometer) at a wavelength of 230 nm against a blank sample^(17, 24, 25).

Evaluation of kinetics data of *in-vitro* drug release

For determination of *in-vitro* release kinetics mechanism of gliclazide floating HBS capsules, the *in-vitro* release data were fitted to mathematically curve fitted kinetics models like zero order, first order, higuchi, korsmeyer peppes and Hickson-crowell model. We observed the n value that will express the release mechanism of drug. If the n value is less than or equal to 0.5

then this is called fickian diffusion controlled release mechanism, if the value ranges between 0.5 to 1 then this is called non-fickian release mechanism and if the value greater than or equal to 1 then it refer to case-II transport or release ^(17, 26).

Evaluation by Fourier Transform-Infrared Spectroscopy (FTIR)

Samples were tested as potassium bromide (IR-grade) pellets by a FTIR spectroscope instrument (Alpha-FTIR, Bruker Optics, Germany). The potassium bromide pellets containing samples were placed in the sample holder, individually. The spectral scanning was recorded over the range of 4000–500 cm^{-1} ^(12, 27).

Evaluated statistical analysis

All determined data are expressed as mean \pm standard deviation (Here n = 5).

RESULTS AND DISCUSSION

Preparation of gliclazide gastroretentive floating HBS cappsules

These HBS capsules of gastroretentive drug delivery showing importance in recent days and specially those are absorbed in small intestine ⁽¹²⁾. This capsules are floated due to the mixing the drug with colloidal gel forming polymer that help to buoyant in the gastric content for long time. In past few decade there are several work reported by taking of chitosan in stomach specific drug delivery due to its gel forming nature as well as in stomach specific or gastroretentive HBS system. Also here I used xanthan gum as a very natural and economical polymer to reliability for compile with chitosan and produce more viscous to hydrate and float for release the drug for long periods of time and also important things in my previous reported work formation of polyelectrolyte complex for sustain release drug delivery between two opposite charge polymer low molecular mass chitosan cationic nature and carboxy methyl tamarind gum anionic nature ^(14, 17, 28-30).

So this HBS capsule of gliclazide will prepared by physical blending of drug gliclazide and two opposite charge hydrophilic polymer chitosan and xanthan gum by adding other excipients like lactose, talc and magnesium stearate as a diluents, glidant and lubricating agent and this blend were encapsulated in a single unit 1 size empty hard gelatin capsule to improve the sustain release of gliclazide from these HBS capsules.

Uniformity of weight

Table 2 will represent the value of weight uniformity of different capsules of gliclazide. All this formulation should follow the USP specification of weight uniformity test. Coefficient of

variation to all formulation not more than 2.50 that's mean all formulation will be filled properly.

Table 2: Weight uniformity test of different floating HBS capsules of gliclazide

Formulation Code	Weight Uniformity	
	Mean Weight (gm)	Coefficient of Variation (%)
GFC1	493.15 ± 4.44	1.01
GFC2	499.65 ± 6.31	1.09
GFC3	494.15 ± 5.67	2.02
GFC4	497.45 ± 4.14	1.05
GFC5	495.20 ± 3.87	1.02

Swelling index

This swelling behaviour is primarily attributed to the inclusion of polyelectrolyte complex formed by two opposite charge polymers, which not only imparts buoyancy to the capsules but also contributes to their expansion. The swelling of the HBS capsules enhances drug release, as it facilitates the diffusion of the drug from the polymer matrix. The swelling percentage was calculated using the specified formula, with the GFC4 formulation exhibiting a swelling percentage of 82%, as shown in Table 3

Drug content uniformity

The value of drug content uniformity is demonstrated in Table 3. The drug content of Gliclazide HBS capsules ranged from 90.68 ± 0.14 to 92.43 ± 0.37 which help to maintain the composition and proper release.

Table 3: Swelling index and drug content uniformity of gastroretentive floating HBS capsule of gliclazide

Formulation Code	Swelling index (%)	Drug content uniformity (%)
GFC1	78.35 ± 0.34	91.05 ± 0.18
GFC2	79.48 ± 0.45	90.68 ± 0.14
GFC3	80.03 ± 0.57	91.03 ± 0.22
GFC4	82.63 ± 0.62	92.43 ± 0.37
GFC5	81.77 ± 0.99	91.87 ± 0.46

In-vitro floatation

The floating timing of all formulations is given in Table 4. It was found that all formulations' was floated not less than 10 hours. These results explain that a significant effect on per cent drug content was observed with polymer concentration.

Table 4: In-vitro floatation of gastroretentive floating HBS capsule of gliclazide

Formulation Code	<i>In-vitro</i> floatation time (Hr)
GFC1	Not less than 10 hours
GFC2	Not less than 10 hours
GFC3	Not less than 10 hours
GFC4	Not less than 10 hours
GFC5	Not less than 10 hours

***In-vitro* drug release**

The *in-vitro* release of gliclazide from these capsules was analyzed in simulated gastric fluid (pH 1.2) showing in Fig 2. Here all the formulation showing good *in-vitro* drug release over a prolong periods of time that more than 10 hours. All these HBS capsules of gliclazide represent well sustain release the drug. It is also notice that the *in-vitro* sustain release of formulation will little bit increase when increase quantity of chitosan by taking appropriate ratio containing anthan gum. Therefore all formulation especially GFC4 showing better sustain release over a prolong periods of time (More than 10 hours). Here another important things to formation of electrolyte complex by taking of two opposite charge polymers i.e. cationic chitosan and

anionic xanthan gum that also improved the sustain release the drug from different formulation.

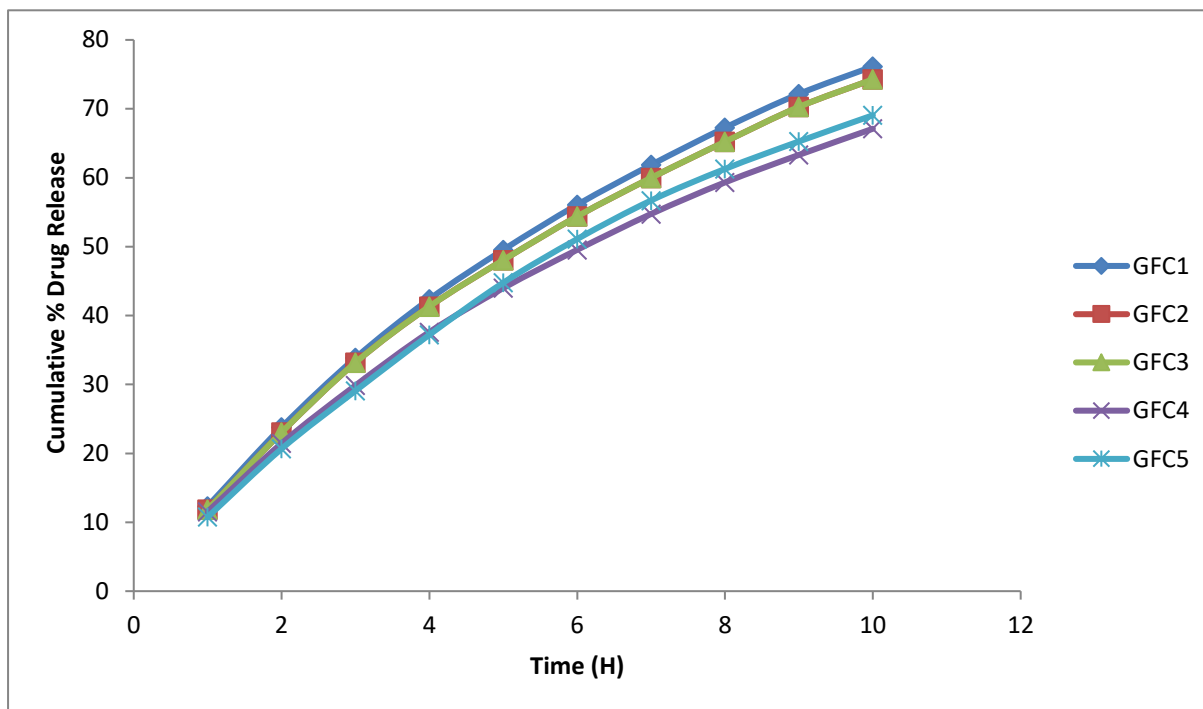


Figure 2: In-vitro release of gliclazide from these floating HBS capsules

In Table 5 represent the different curve fitting mathematical kinetics model. All the formulation of gliclazide floating HBS capsule will fitted in Korsmeyer-Peppas model ($R^2 = 0.991$ to 0.998). The release exponent (n) of all formulation was showing ranges within 0.77 to 0.79 refer to release mechanism like non-Fickian or anomalous diffusion indicating both controlled swelling and diffusion.

Table 5: In-vitro release kinetics data of all formulation

Formulation Code	R ² Value					n value
	Zero order	First order	Higuchi	Korsmeyer-peppes	Hickson-crowell	
GFC1	0.975	0.845	0.877	0.991	0.902	0.77
GFC2	0.974	0.843	0.876	0.998	0.901	0.78
GFC3	0.974	0.848	0.874	0.992	0.903	0.77
GFC4	0.975	0.853	0.889	0.993	0.906	0.79
GFC5	0.976	0.852	0.888	0.994	0.907	0.78

FTIR characterization

FTIR spectra of gliclazide, chitosan, xanthan gum and formulation GFC4 for determination of drug-polymer interaction will be showing on Fig. 3. The FTIR spectrum of this formulation

will show different characteristic peaks due to the stretching and bending of presence of different functional group. This all type of peaks compare to the drug and polymers that there is no more interaction as well as chemical or physical incompatibilities occur in between drug-polymer which maintain the stability.

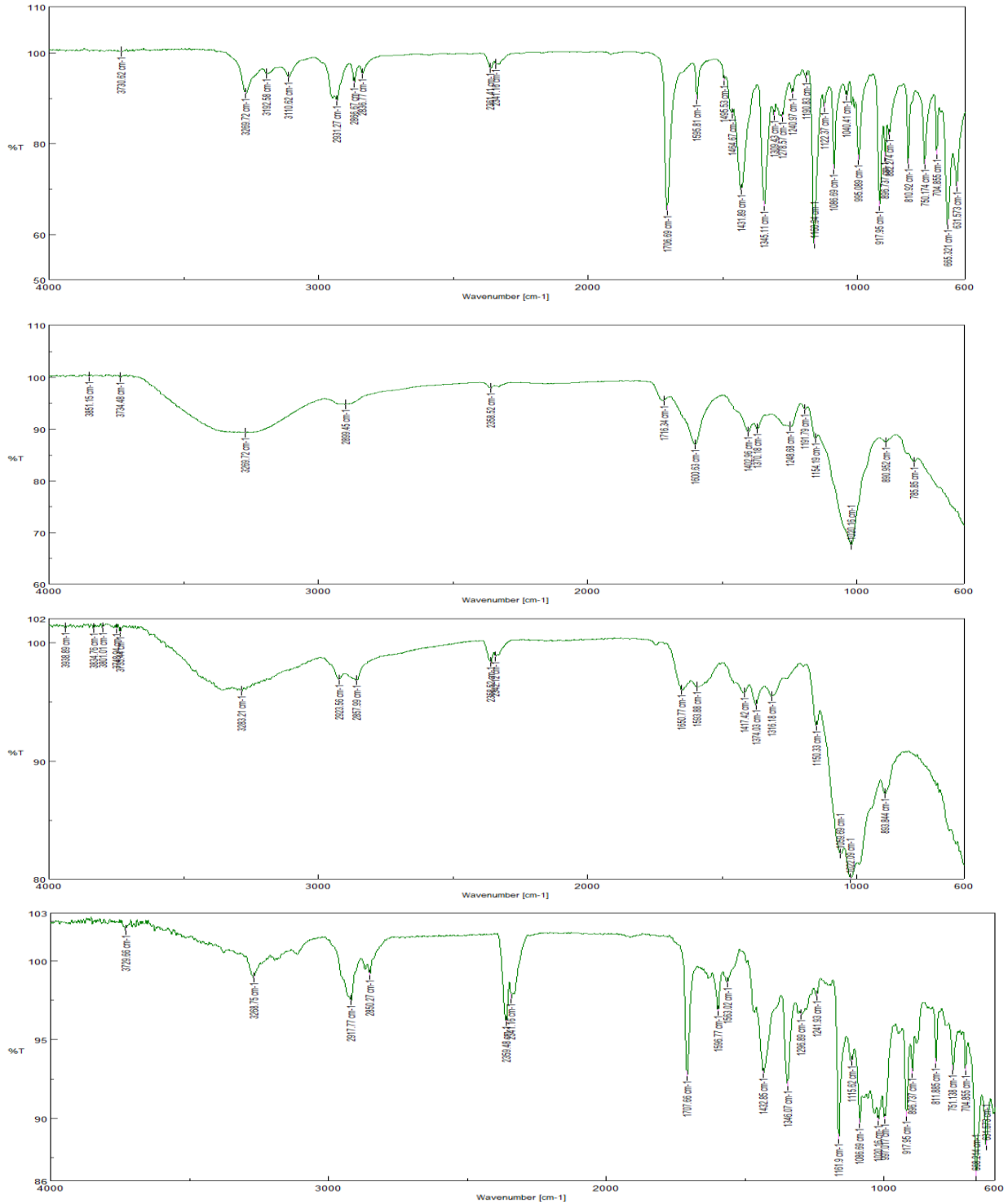


Figure 3: FTIR spectra of gliclazide, chitosan, xanthan gum and GFC4 HBS capsule

CONCLUSION

From this research work it was concluded that the formulation of gastroretentive floating HBS capsules of gliclazide prepared by physical blending of two opposite charge hydrophilic gel forming colloidal polymer, namely cationic chitosan and anionic xanthan gum (formation of electrolyte complex) help to sustain release of drug. The different evaluated properties like uniformity of weight, drug content uniformity, swelling activity, *in-vitro* floatation prove that good drug present due to better mixing, excellent swelling and long floating retention to maintain the stability and developed the system. All these formulation showing great sustain drug release (*in-vitro*) in simulated gastric fluid (pH: - 1.2) over more than 10 hours, although the GFC4 showing best sustain among this. This is because proper the ratio of two opposite charge polymer and to form the polyelectrolyte complex. So this type of gastroretentive floating HBS capsules will be very useful in the future to other suitable characterized drugs which help to retain and sustain release the drug over prolong periods of time.

ACKNOWLEDGEMENTS

The authors express their sincere gratitude to Kalinga Institute of Pharmaceutical Sciences, Biruan, Balasore, Odisha, India, and the respective affiliated institutions for providing the necessary laboratory facilities, infrastructure, and academic support to carry out this research work successfully.

AUTHOR CONTRIBUTIONS

Radheshyam Samanta: Conceptualization, formulation development, experimental work, data analysis, interpretation of results, and manuscript preparation.

Tamal Deb Goswami: Literature review, experimental assistance, data compilation, and manuscript editing.

Hrushikesh Behera: Formulation optimization, characterization studies, and data collection.

Ranjit Kumar Sutar: In-vitro evaluation studies, statistical analysis, and result interpretation.

Ashok Kumar Sethi: Research supervision, methodology validation, and critical review of the manuscript.

Pravati Deo: Scientific guidance, manuscript review, and technical support.

Bhabatarini Jena: Final manuscript review, proofreading, and approval of the final version.

All authors have read and approved the final manuscript.

FUNDING

No external funding was received for conducting this research study.

CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest regarding the publication of this manuscript.

DATA AVAILABILITY STATEMENT

The data supporting the findings of this study are available from the corresponding author upon reasonable request.

CORRESPONDING AUTHOR

Dr. Radheshyam Samanta

Department of Pharmaceutics,
Kalinga Institute of Pharmaceutical Sciences,
Biruan, Balasore – 756060, Odisha, India.
Email: radheshyamsamanta93@gmail.com

REFERENCE

1. Arrora, S., Ali, J., Khar, R.K., Baboota, S., (2005). Floating drug delivery systems: A review. AAPS PharmSciTech 6(3): 372-390.
2. Borase, C.B., Floating system for oral controlled release drug delivery. A Review. International Journal of Applied Pharmaceutics. 2012; 4(2): 1-13.
3. Bhowmik, D., Chiranjib, B., Chandira, M., Jayakar, B., Kumar S. Floating drug delivery system: A Review. Scholars Research Library Der Pharmacia Lettre. 2009; 1(2): 199 - 218.
4. Dhiman S, Philip N, Gurjeet Singh T, Babbar R, Garg N, Diwan V. An insight on novel approaches and perspectives for gastro-retentive drug delivery systems. Curr Drug Deliv. 2023;20(6):708-29.
5. Mohapatra PK, Satyavani CH, Sahoo S. The design and development of carvedilol gastroretentive floating drug delivery systems using hydrophilic polymers and *in-vitro* characterization. Int J Pharm Pharm Sci. 2020;12(7):66-73. doi: 10.22159/ijpps.2020v12i7.38024.

6. Juthi AZ, Li F, Wang B, Alam MM, Talukder ME, Qiu B. pH-responsive super-porous hybrid hydrogels for gastroretentive controlled-release drug delivery. *Pharmaceutics*. 2023;15(3):816.
7. S. Dhiman, N. Philip, T. Gurjeet Singh, R. Babbar, N. Garg, V. Diwan, P. Singh, An insight on novel approaches & perspectives for gastro-retentive drug delivery systems, *Curr. Drug Deliv*. 20 (2023) 708–729
8. Sahni, J.K., Ahmad, F.J., Ahuja, A., Khar, R.K., Formulation and evaluation of a hydrodynamically balanced system of paracetamol. *The Indian Pharmacist* 2006; V (46): 64-66.
9. Ali, J., Hasan S, Ali, M., (2006). Formulation and development of gastroretentive drug delivery system for ofloxacin. *Methods Find Exp Clin Pharmacol* 28: 1–7.
10. Wason, R., Nanda, A., Development and optimization of a gastroretentive FDDS using ciprofloxacin hydrochloride. *The Indian Pharmacist* 2009; Dec: 81-89.
11. Dorozynski, P., Kulinowski, P., Jachowicz, R., et al. (2007). Development of a simultaneous studies and magnetic resonance imaging of water transport in hydrodynamically balanced systems: A technical note. *AAPS PharmSciTech* 8, Article 15.
12. Samanta, R, Nayak, S, Das, B, Nayak, A.K., Chitosan-carboxymethyl tamarind gum *in situ* polyelectrolyte complex-based floating capsules of ofloxacin: *In-vitro-in vivo studies*, *International Journal of Biological Macromolecules*, 253 (2023) 127507.
13. L. Raju, Anu Sharma, Abhishek Soni. Formulation and Evaluation of Glimepiride Floating Beads. *WJPRT*, 2015, Vol. 3(3).
14. Verma, A, Dubey, J, Verma N, and Nayak, A.K., Chitosan-Hydroxypropyl Methylcellulose Matrices as Carriers for Hydrodynamically Balanced Capsules of Moxifloxacin HCl, *Current Drug Delivery*, 2016, Vol. 13, No. 0.
15. M. Nafady, K. Attallah, M. Sayed, A. Gouda. Formulation and Evaluation of a Buoyant Ranitidine Hydrochloride System. *Int. J. Pharm. Sci. Rev. Res.*, 24(2), Jan – Feb 2014; n° 02, 4-8.
16. Venkateswarlu, k, Jami K.P., Badithala S.S.K., Formulation Development and In-vitro Evaluation of Floating Tablets of Ciprofloxacin HCl. *Asian Journal of Pharmaceutics* Oct-Dec 2016 10 (4) | 271.

17. Samanta, R, Tiwari, G, Gupta, N, Rajput, D.S., Designing, development and evaluation of gastroretentive floating hbs system of metformin: *in-vitro in vivo* studies. Int J App Pharm, Vol 16, Issue 5, 2024, 258-265.
18. Janaa, S, Samanta A, Nayak, A.K., Sena, K.K., Jena, Subrat., Novel alginate hydrogel core–shell systems for combination delivery of ranitidine HCl and aceclofenac. International Journal of Biological Macromolecules 74 (2015) 85–92.
19. Raju DB, Sreenivas R, Varma MM. Formulation and evaluation of floating drug delivery system of metformin hydrochloride. J Chem Res. 2010;2(2):274-8.
20. Verma A, Bansal AK, Ghosh A, Pandit JK. Low molecular mass chitosan as carrier for a hydrodynamically balanced system for sustained delivery of ciprofloxacin hydrochloride. Acta Pharm. 2012;62(2):237-50.
21. Verma A, Dubey J, Verma N, Nayak AK. Chitosan-hydroxypropyl methylcellulose matrices as carriers for hydrodynamically balanced capsules of moxifloxacin HCl. Curr Drug Deliv. 2017;14(1):83-90.
22. Malakar J, Datta PK, Purakayastha SD, Dey S, Nayak AK. Floating capsules containing alginate-based beads of salbutamol sulfate: *in-vitro-in vivo* evaluations. Int J Biol Macromol. 2014;64:181-9.
23. E.A. Klausner, E. Lavy, D. Stepensky, M. Friedman, A. Hoffman, Novel gastro-retentive dosage forms: evaluation of gastroretentivity and its effect on riboflavin absorption in dogs, Pharm. Res. 19 (2002) 1516–1523.
24. A.K. Nayak, B. Das, R. Maji, Gastroretentive hydrodynamically balanced system of ofloxacin: formulation and *in-vitro* evaluation, Saudi Pharm. J. 21 (2013) 113–117.
25. Javed A, Shweta A, Formulation and development of hydrodynamically balanced system for metformin: *in-vitro* and *in vivo* evaluation, Euro J Pharm Biopharm, 67, 2007, 196-201.
26. J. Ali, S. Arora, A. Ahuja, A.K. Babbar, R.K. Sharma, R.K. Khar, S. Baboota, Formulation and development of hydrodynamically balanced system for metformin: *in-vitro* and *in vivo* evaluation, Eur. J. Pharm. Biopharm. 67 (2007) 196–201.
27. Bopanna R, Kulkarni RV, Setty CM. Carboxymethyl cellulose-aluminium hydrogel microbeads for prolong release of Simvastatin. Acta Pharmaceutica Scientia. 2010; 52(2): 137-143.

28. A. Verma, A.K. Bansal, A. Ghosh, J.K. Pandit, Low molecular mass chitosan as carrier for a hydrodynamically balanced system for sustained delivery of ciprofloxacin hydrochloride, Acta Pharm. 62 (2012) 237–250.
29. Ray S, Banerjee S, Maiti S, Laha B, Barik S, Sa B. Novel interpenetrating network microspheres of xanthan gum-poly(vinyl alcohol) for the delivery of diclofenac sodium to the intestine—*in-vitro* and *in vivo* evaluation. Drug Deliv. 2010;17(7):508-19.
30. Reddy MS, Begum Z. Formulation and *in-vitro* evaluation of gastroretentive in situ floating gels of telmisarten cubosomes. Int J Curr Pharm Sci. 2022;14(1):44-53.